

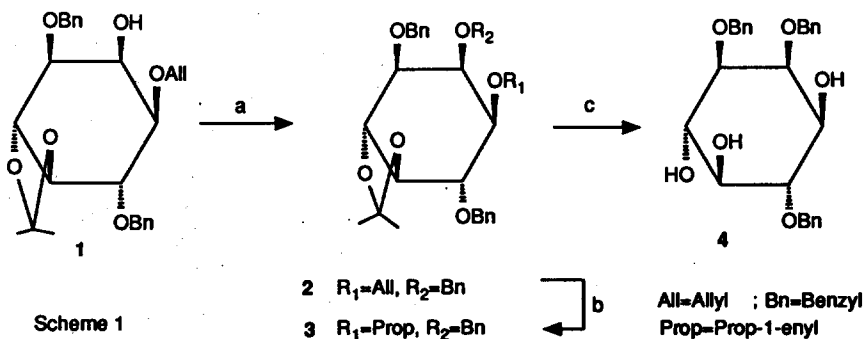
SYNTHESIS OF 1,4,5-TRISSULPHATED AND 1,4,5-TRISSULPHAMOYLATED *myo*-INOSITOLS: ISOSTERIC *myo*-INOSITOL 1,4,5-TRISPHOSPHATE ANALOGUES

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Abstract: *Synthesis of the myo-inositol 1,4,5-trisphosphate isosteric analogues (rac.) 1,4,5-trissulphate and (rac.) 1,4,5-trissulphonamido was accomplished from 1,2,4-tri-O-benzyl myo-inositol.*

It is now well understood that D-*myo*-inositol 1,4,5-trisphosphate (IP₃), generated by a receptor mediated hydrolysis of a phosphoinositide, acts as an intracellular second messenger¹. IP₃ has been shown to release Ca⁺⁺ from intracellular stores, initiating a number of physiological responses. IP₃ is metabolized via two distinct pathways: IP₃ dephosphorylation by a 5-phosphatase gives inositol 1,4-bisphosphate, whereas phosphorylation by a 3-kinase yields another putative second messenger, inositol 1,3,4,5-tetraphosphate², the potential role of which is currently under investigation.

Since IP₃ is still the only inositol phosphate for which a clear-cut role in intracellular Ca⁺⁺ homeostasis has been demonstrated, there is a considerable interest in the synthesis³ of IP₃ and derivatives thereof. In order to design IP₃ derivatives acting as intracellular Ca⁺⁺-antagonist, a better understanding of the mechanism of Ca⁺⁺ release at a molecular level will be essential. Mapping of the substrate specificity of the IP₃ receptor proteins can be performed when systematically modified analogues of the natural ligand (i.e. IP₃) are available. Recent advances indicated the vicinal phosphate groups⁴ at position 4,5 and the 3- and 6-hydroxyl groups⁵ to be essential for Ca⁺⁺ release. Additional structure activity relationships can be

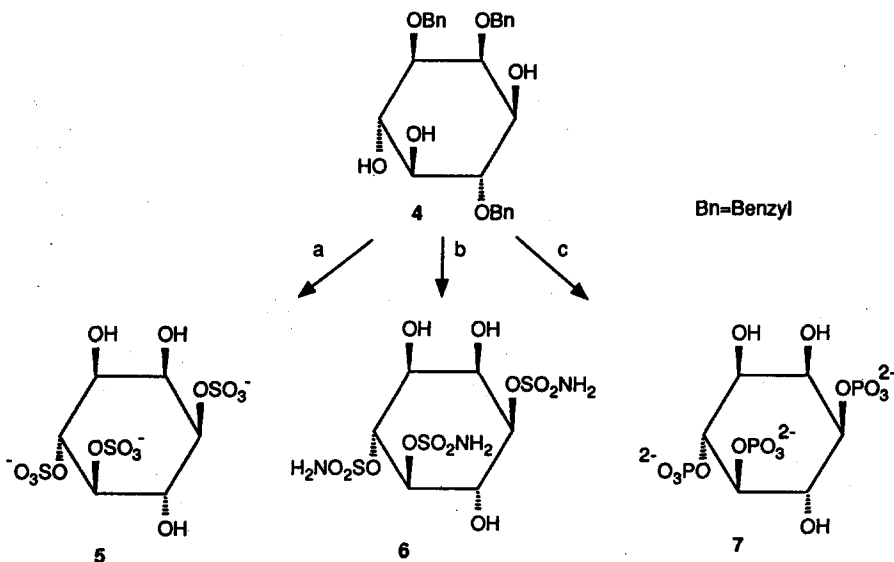


a: NaH/BnBr/DMF; b: Ir[COD(PMePh₂)₂]PF₆/H₂; c: HCl/dioxane/MeOH

gathered from IP₃ analogues containing phosphate modified functions or phosphate isosteric groups. Apart from the synthesis of derivatives containing phosphorothioate moieties⁶, no other report on the synthesis of isosteric IP₃ analogues has been published so far.

Since it has previously been shown that the sulphated polysaccharide, heparin, may compete with IP₃ for its receptor binding⁷, the synthesis and biological testing of myo-inositol 1,4,5-trissulphate will be of considerable interest. It is not yet clear, however, whether the affinity of heparin for the IP₃ receptor can be attributed to a binding of sulphate groups in regions normally accommodating the IP₃ phosphates. The synthesis of neutral and isosteric analogues will be feasible by applying sulphonamide groups. In this communication, we wish to report the synthesis of two isosteric analogues of myo-inositol 1,4,5-trisphosphate, i.e. myo-inositol 1,4,5-trissulphate **5** and myo-inositol 1,4,5-trissulphonamide **6**.

Properly protected **1** was obtained from myo-inositol by the literature procedure⁸. Regioselective stannylene-mediated allylation⁹ of **1** and subsequent benzylation afforded fully protected **2** (73% yield, see Scheme 1). Treatment of **2** with cyclooctadiene-bis(methyldiphenylphosphine)-iridium hexafluorophosphate¹⁰ afforded **3** (100% yield), which on acid hydrolysis (dioxane/methanol/0.1 N HCl) furnished key-intermediate triol **4**³ (96% yield).



Scheme 2

*a: (CH₃CH₂)₃N.SO₃/DMF then 10% Pd on C/H₂/DMF/H₂O; b: NH₂SO₂Cl/NaH/DMF then 10% Pd on C/H₂ in DMF/H₂O; c: (CNCH₂CH₂O)₂PN(CH₂CH₃)₂/1-H-tetrazole in CH₂Cl₂/CH₃CN followed by *tert*-BuOOH/Et₃N and 0.2N NaOH/dioxane/MeOH; 10% Pd on C/H₂/DMF/H₂O.*

We now turned our attention to the introduction of the different functional groups, i. e. sulphate and sulphonamide (Scheme 2). Treatment of **4** (0.2 mmol) for 16 hr at 50°C with triethylamine sulphur trioxide complex¹¹ (3 mmol) in DMF provided a protected sulphated inositol intermediate (R_f 0.35, EtOAc: Pyr: AcOH: H₂O, 11: 7: 1.6: 4). The latter was purified by Sephadex LH-20 column chromatography¹² (eluent DMF containing 0.5% triethylamine) and silica gel column chromatography

(eluent: $\text{CH}_2\text{Cl}_2/\text{MeOH}$, 4:1). Subsequent hydrogenolysis (10% Pd on C in 4:1 DMF/ H_2O) furnished myo-inositol 1,4,5-trissulphate **5** in a yield of 86% from **4**.

In order to obtain the neutral 1,4,5-trissulphamoylated derivative **6**, triol **4** (0.2 mmol) was treated with sulphamoyl chloride¹³ (1.2 mmol) and NaH in DMF at 0°C for 2 hr to give, after silica gel column chromatography (eluent: $\text{CH}_2\text{Cl}_2/\text{MeOH}$, 95:5), the protected intermediate (R_f 0.6, $\text{CH}_2\text{Cl}_2/\text{MeOH}$, 99:1). Subsequent debenylation (10% Pd on C in DMF/ H_2O) gave inositol 1,4,5-trissulphonamide **6** (R_f 0.25, RP18 silica, EtOH/ H_2O , 3:1) in 84% yield from **4**. Starting from compound **4** we also synthesized (*rac.*) IP_3 itself (**7**), as a reference compound, by slight modification of literature procedures¹⁴.

The identity and homogeneity of the compounds **5,6** and **7** were established by NMR spectroscopy and (FAB) mass spectrometry. The $^1\text{H-NMR}$ spectral data are listed in the Table¹⁶. The biological data on the title compounds will be reported elsewhere in due time.

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12. The combined appropriate fractions were treated with NaHCO_3 (1 eq./ OSO_3^-) prior to concentration.
13. Sulphamoyl chloride was prepared from chlorosulphonyl isocyanate by controlled hydrolysis.
14. Treatment of triol 4 at 0° – 20°C for 1 hr with bis(2-cyanoethyl)-*N,N*-diethyl-phosphoramidite¹⁵ in the presence of 1*H*-tetrazole followed by oxidation for 4 hr with excess *tert*-butyl hydroperoxide in the presence of triethylamine afforded a phosphodiester intermediate, which was subjected to Sephadex LH-20 column chromatography. Base treatment (0.2 N NaOH) of the appropriate fractions and subsequent hydrogenolysis (10% Pd on C in *tert*-butanol/ H_2O for 16 hr) afforded crude 7, which was applied to DEAE column chromatography ($1 \times 30 \text{ cm}^2$, 0.3–1.0 M NH_4OAc) to give 7 in 76% overall yield.
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- 16.

inositol 1,4,5-	H_1	H_2	H_3	H_4	H_5	H_6
trissulphate (5)	4.27 dd	4.41 t	3.85 dd	4.52 t	4.29 t	3.96 t
trissulphomamide (6)	4.55 dd	4.57 t	3.97 dd	4.79 t	4.64 t	4.16 t
triphosphate (7)	4.10 c	4.26 c	3.71 dd	4.28 c	4.10 c	3.88 t

Table, $^1\text{H-NMR}$ chemical shifts (360 MHz) of compounds 5, 6 and 7.

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